



AccuPrime™ *Pfx* DNA Polymerase

Cat. No. 12344-024
12344-032

Size: 200 Reactions
1000 Reactions

Conc. 2.5 U/μl

Store at -20°C

Description

AccuPrime™ *Pfx* DNA Polymerase is a proprietary enzyme preparation containing recombinant DNA polymerase from *Thermococcus* species strain KOD (1,2). This polymerase possesses a proofreading 3' to 5' exonuclease activity that provides higher fidelity than *Pfu* DNA polymerase (3).

AccuPrime™ *Pfx* DNA Polymerase is a highly processive enzyme and possesses a fast chain extension capability. It is provided in an antibody-bound form that is inactive at ambient temperatures. The enzyme regains activity after the initial denaturation step at 94°C in PCR cycling, providing an automatic "hot start" that increases specificity, sensitivity, and yield, while allowing room temperature assembly (4).

10X AccuPrime™ *Pfx* Reaction Mix contains thermostable AccuPrime™ proteins, MgSO₄, and dNTPs. Thermostable AccuPrime™ proteins enhance specific primer-template hybridization during every cycle of PCR (5). The high specificity, fidelity, and yield offered by AccuPrime™ *Pfx* DNA Polymerase make it ideal for demanding PCR applications such as site-directed mutagenesis and PCR expression cloning.

Reagents are provided for 200 or 1000 amplification reactions of 50 μl each.

<u>Component</u>	<u>200-Rxn kit</u>	<u>1000-Rxn kit</u>
AccuPrime™ <i>Pfx</i> DNA Polymerase (2.5 U/μl)	100 μl	500 μl
50-mM Magnesium Sulfate	1 ml	2 × 1 ml
10X AccuPrime™ <i>Pfx</i> Reaction Mix	1 ml	5 × 1 ml

Unit Definition

One unit of AccuPrime™ *Pfx* DNA Polymerase incorporates 10 nmol of deoxyribonucleotide into acid-insoluble material in 30 min at 74°C.

Part. no. 12344.pps

Rev. date: 07/11/03

This product is distributed for laboratory research only. CAUTION: Not for diagnostic use. The safety and efficacy of this product in diagnostic or other clinical uses has not been established.

AccuPrime™ Pfx DNA Polymerase Storage Buffer

50-mM Tris-HCl (pH 8.0), 50-mM KCl, 1-mM DTT, 0.1-mM EDTA, stabilizers, and 50% (v/v) glycerol

Quality Control

AccuPrime™ Pfx DNA Polymerase is functionally tested in an amplification reaction using 100 ng of K562 genomic DNA. A DNA polymerization activity assay measures percent of DNA polymerase inhibition versus an uninhibited control. AccuPrime™ proteins are tested for absence of double- and single-strand endonuclease activity and absence of 5' and 3' exonuclease activity.

General Recommendations and Guidelines for PCR

PCR is a powerful technique capable of amplifying trace amounts of DNA. All appropriate precautions should be taken to avoid cross-contamination.

MgSO₄: MgSO₄ is included in the 10X AccuPrime™ Pfx Reaction Mix at a final concentration of 1 mM, which is sufficient for most templates. For further optimization, add 0.1 µl to 1.0 µl of 50-mM MgSO₄ (included in the kit) to the reaction.

dNTPs: dNTPs are included in the 10X AccuPrime™ Pfx Reaction Mix at a final concentration of 0.3 mM.

Annealing Temperature: The optimal annealing temperature should be 5–10°C lower than the T_m of the primers used; if necessary, gradually increase the annealing temperature by 2–3°C for higher specificity.

KCl: For difficult primer sets, prepare titrations of KCl (not included) at final concentrations of 20–50 mM for further optimization.

PCR Protocol

The following general procedure is suggested as a starting point when using AccuPrime™ *Pfx* DNA Polymerase in any PCR amplification.

1. Add the following components to an autoclaved microcentrifuge tube at either room temperature or on ice:

<u>Component</u>	<u>Volume</u>	<u>Final Conc.</u>
10X AccuPrime™ <i>Pfx</i> Reaction mix*	5 µl	1X
Primer mix (10 µM each)*	1.5 µl	0.3 µM each
Template DNA (10 pg–200 ng)	≥1 µl	As required
AccuPrime™ <i>Pfx</i> DNA Polymerase**	0.4–1 µl	1.0–2.5 units
Autoclaved, distilled water	to 50 µl	

*AccuPrime™ *Pfx* DNA Polymerase will not function in reactions that contain dUTP either in the primers or in the dNTP mix.

**For most targets, 1 unit is optimal. Higher concentrations may be inhibitory. More enzyme may be required for longer targets (>3 kb).

2. Mix contents of the tubes and overlay with mineral or silicone oil, if necessary. (Note: The oil overlay is unnecessary in thermal cyclers equipped with a heated lid.)
3. Cap the tubes and centrifuge briefly to collect the contents.
4. Denature the template for 2 min at 95°C. Perform 25–35 cycles of PCR amplification as follows:

<u>Three-step cycling</u>	<u>Two-step cycling</u>
Denature: 95°C for 15 s	Denature: 95°C for 15 s
Anneal: 55–64°C for 30 s	Extend: 68°C for 1 min per kb
Extend: 68°C for 1 min per kb	

Note: Two-step cycling can be used for long primers with high T_m .

5. Maintain the reaction at 4°C after cycling. The samples can be stored at -20°C until use.
6. Analyze the products by agarose gel electrophoresis and visualize by ethidium bromide staining.

References

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2. Nishioka M, Mizuguchi H, Fujiwara S, Komatsubara S, Kitabayashi M, Uemura H, Takagi M, Imanaka T. (2001) *J. Biotechnol.*, **88**, 141-9.
3. Cline, J., Braman, and Hogrefe, H. H. (1996) *Nucleic Acid Res.*, **24**, 3546.
4. Sharkey, D.J., Scalice, E.R., Christy, K.G., Atwood, S.M., Daiss, J.L. (1994) *BioTechnology*, **12**, 506.
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