Assay Designs, Inc.

TiterZyme ® EIA

rat GRO/CINC-1

Enzyme Immunometric Assay Kit

Catalog No. 900-074 (formerly 90074)

96 Determination Kit

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Description

Assay Designs' rat GRO/CINC-1 (rat IL-8) TiterZyme® Enzyme Immunometric Assay (EIA) kit is a complete kit for the quantitative determination of rat GRO/CINC-1 in biological fluids. Please read the complete kit insert before performing this assay. The kit uses a polyclonal antibody to rat GRO/CINC-1 immobilized on a microtiter plate to bind the rat GRO/CINC-1 in the standards or sample. A recombinant rat GRO/CINC-1 Standard is provided in the kit. After a short incubation the excess sample or standard is washed out and a polyclonal antibody to rat GRO/CINC-1 labeled with the enzyme Horseradish peroxidase is added. This labeled antibody binds to the rat GRO/CINC-1 captured on the plate. After a short incubation the excess labeled antibody is washed out and substrate is added. The substrate reacts with the labeled antibody bound to the rat GRO/CINC-1 captured on the plate. After a short incubation, the enzyme reaction is stopped and the color generated is read at 450 nm. The measured optical density is directly proportional to the concentration of rat GRO/CINC-1 in either standards or samples. For further explanation of the principles and practice of immunoassays please see the excellent books by Chard[1] or Tijssen[2].

Introduction

Cytokine-induced Neutrophil Chemoattractant-1 (CINC-1) was originally purified from media conditioned by IL-1β stimulated rat kidney epithelioid cells (NRK-52E)[3-7]. Watanabe's group at Toyoma Medical and Pharmaceutical University identified the amino acid sequence that encodes for rat CINC-1 in 1989. CINC-1 is a member of the alpha (CXC) subfamily of chemokines. Three additional rat CXC chemokines (CINC-2α, CINC-2β, CINC-3/MIP-2) have been identified. The protein sequence of CINC-1 is 63-67% identical to that of CINC-2α, CINC-2β, and CINC-3/MIP-2. In addition the Growth Related Oncogene (GRO), GROα, GROβ and GROγ shares 68%, 71% and 69% identity with CINC-1. It has been suggested that CINCs are the rat counter-part of human GROs.

Precautions

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1. Stop Solution is a 1 normal (1N) sulfuric acid solution. This solution is caustic; care should be taken in use.
2. The activity of the Horseradish peroxidase conjugate is affected by nucleophiles, such as azide, cyanide and hydroxylamine.
3. We test this kit's performance with a variety of samples, however it is possible that high levels of interfering substances may cause variation in assay results.
4. The rat GRO/CINC-1 Standard provided, Catalog No. 80-0676, should be handled with care, because of the known and unknown effects of GRO/CINC-1.
5. The rat GRO/CINC-1 Standard and Labeled Antibody should be stored at -20˚C. Do not repeatedly freeze/thaw.

Materials Supplied

1. **rat GRO/CINC-1 Microtiter Plate, One Plate of 96 Wells, Catalog No. 80-0674**
   A strip microtiter plate coated with rabbit antibody specific to rat GRO/CINC-1.
2. **rat GRO/CINC-1 Labeled Antibody, 1 vial, Catalog No. 80-0675**
   Rabbit antibody to rat GRO/CINC-1 conjugated to Horseradish peroxidase.
3. **Assay Buffer, 30 mL, Catalog No. 80-0170**
   Phosphate buffered saline containing proteins and detergents.
4. **Labeled Antibody Diluent, 10 mL, Catalog No. 80-0182**
   Phosphate buffered saline containing proteins and detergents.
5. **Wash Buffer Concentrate, 50 mL, Catalog No. 80-0171**
   Phosphate buffered saline containing detergents.
6. **rat GRO/CINC-1 Standard, 1 vial, Catalog No. 80-0676**
   A vial containing 300 pg of recombinant rat GRO/CINC-1.
7. **Substrate Buffer, 5 mL, Catalog No. 80-0173**
   A solution of phosphate in buffer. Ready to use.
8. **Peroxide Solution, 5.5 mL, Catalog No. 80-0174**
   A 0.01% solution of hydrogen peroxide in water. Ready to use.
9. **TMB Tablets, 2 Tablets, Catalog No. 80-0175**
10. **Stop Solution, 11 mL, Catalog No. 80-0176**
    A 1N solution of sulfuric acid in water. Keep tightly capped. Caution: **Caustic.**
11. **rat GRO/CINC-1 Assay Layout Sheet, 1 each, Catalog No. 30-0139**
12. **Plate Sealer, 2 each, Catalog No. 30-0012**

Storage

All components of this kit, **except the Labeled Antibody and Standard**, are stable at 4ºC until the kit's expiration date. The Labeled Antibody and Standard **must** be stored at -20ºC.

Materials Needed but Not Supplied

1. Deionized or distilled water.
2. Precision pipets for volumes between 100 µL and 1,000 µL.
3. Disposable test tubes for dilution of samples and standards.
4. Repeater pipet for dispensing 100 µL.
5. Disposable beakers for diluting buffer concentrates.
6. Graduated cylinders.
7. A 37°C incubator.
8. Adsorbent paper for blotting.
9. Microplate reader capable of reading at 450 nm., preferably with correction between 570 nm and 590 nm.
10. Graph paper for plotting the standard curve.

Sample Handling

Assay Designs' TiterZyme® EIA is compatible with rat GRO/CINC-1 (rat IL-8) samples in a wide range of matrices. Samples diluted sufficiently into Assay Buffer can be read directly from the standard curve. Please refer to the Sample Recovery recommendations on page 11 for details of suggested dilutions.

Culture fluids and serum are suitable for use in the assay. Samples containing visible precipitate should be clarified prior to use in the assay. Do not use grossly hemolyzed or lipemic specimens. Samples in the majority of tissue culture media, including those containing fetal bovine serum, can also be read in the assay if diluted into Assay Buffer. Users should only use standard curves generated in Assay Buffer to calculate concentrations of rat GRO/CINC-1.

Procedural Notes

1. Do not mix reagents from different kit lots or use reagents beyond the kit expiration date.
2. Allow all reagents to warm to room temperature for at least 30 minutes before opening.
3. Standards can be made up in either glass or plastic tubes.
4. Pre-rinse the pipet tip with reagent, use fresh pipet tips for each sample, standard and reagent.
5. Pipet standards and samples to the bottom of the wells.
6. Add the reagents to the side of the well to avoid contamination.
7. This kit uses plates with removable strips. Unused strips must be kept desiccated at 4ºC in the sealed foil bag. The strips should be used in the frame provided.
8. **Prior to addition of standard, antibody, and substrate, ensure that there is no residual wash buffer in these wells. Any remaining wash buffer may cause variation in assay results.**

Reagent Preparation

1. Wash Buffer

Prepare Wash Buffer by diluting 25 mL of the supplied concentrate with 975 mL of deionized water. This can be stored at 4ºC until the kit expiration date, or for 3 months, whichever is earlier.

2. rat GRO/CINC-1 Standards

Add 500 µL of deionized water to the rat GRO/CINC-1 Standard. Let it sit at room temperature for 5 minutes. Mix it gently. This solution contains 600 pg/mL rat GRO/CINC-1.

Label seven 12 x 75 mm glass tubes #1 through 7. Pipet 230 µL of Assay Buffer into tubes #1 through #7. Add 230 µL of the 600 pg/mL standard to tube #1. Vortex. Add 230 µL of tube #1 to tube #2 and vortex thoroughly. Continue this for tubes #3 through #7.

The concentration of rat GRO/CINC-1 in tubes #1 through #7 will be 300, 150, 75, 37.5, 18.8, 9.38, and 4.69 pg/mL respectively. See rat GRO/CINC-1 Assay Layout Sheet for dilution details. STORE STANDARD AT -20˚C, avoid repeated freeze/thaws.

3. Preparation of Labeled Antibody Conjugate

Add the entire contents of one (1) bottle of Labeled Antibody Diluent to the vial of rat GRO/CINC-1 Antibody Conjugate. Let it stand at room temperature for 5 minutes and then vortex it gently. Use within 15 minutes. After reconstitution, any unused Labeled Antibody should be aliquoted and stored at -20ºC.

4. Preparation of Substrate

Just prior to addition of the substrate solution to the plate, prepare the substrate by adding 1 substrate tablet to 2.5 mL of Substrate Buffer and mix allowing the tablet to completely dissolve before proceeding. Ensure that the tablet has completely dissolved before proceeding. Add 2.75 mL of the Peroxide Solution to this and mix well. Use within 15 minutes.

Assay Procedure

Bring all reagents to room temperature for at least 30 minutes prior to opening.

All standards and samples should be run in duplicate.

1. Refer to the Assay Layout Sheet to determine the number of wells to be used and put any remaining wells with the desiccant back into the foil pouch and seal the ziploc. Store unused wells at 4°C.

2. Pipet 100 µL of Assay Buffer into the S0 (0 pg/mL Standard) wells.

3. Pipet 100 µL of Standards #1 through #7 into the appropriate wells.

4. Pipet 100 µL of the Samples into the appropriate wells.

5. Tap the plate gently to mix the contents.

6. Seal the plate and incubate at 37°C for 1 hour.

7. Empty the contents of the wells and wash by adding 400 µL of wash solution to every well. Repeat the wash 9 more times for a total of **10 washes**. After the final wash, empty or aspirate the wells, and firmly tap the plate on a lint free paper towel to remove any remaining wash buffer.

8. Pipet 100 µL of the Labeled Antibody into each well, except the Blank.

9. Seal the plate and incubate at 37ºC for 30 minutes. Prepare Substrate (See page 5, Section 4).

10. Empty the contents of the wells and wash by adding 400 µL of wash solution to every well. Repeat the wash 11 more times for a total of **12 washes**. After the final wash, empty or aspirate the wells, and firmly tap the plate on a lint free paper towel to remove any remaining wash buffer.

11. Add 100 µL of the Substrate Solution to each well.

12. Incubate for 30 minutes at room temperature in the dark.

13. Add 100 µL of Stop Solution to each well.

14. Blank the plate reader against the Blank wells, read the optical density at 450nm, preferably with correction between 570 and 590 nm. If the plate reader is not able to be blanked against the Blank wells, manually subtract the mean optical density of the blank wells from all readings.

Calculation of Results

Several options are available for the calculation of the concentration of rat GRO/CINC-1 in the samples. We recommend that the data be handled by an immunoassay software package utilizing a 4 parameter logistic curve fitting program. If data reduction software is not readily available, the concentration of rat GRO/CINC-1 can be calculated as follows:

1. Calculate the average net Optical Density (OD) bound for each standard and sample by subtracting the average Blank OD from the average OD for each standard and sample.

   Average Net OD = Average OD - Average Blank OD

2. Using linear graph paper, plot the Average Net OD for each standard versus rat GRO/CINC-1 concentration in each standard. Approximate a straight line through the points. The concentration of rat GRO/CINC-1 in the unknowns can be determined by interpolation.

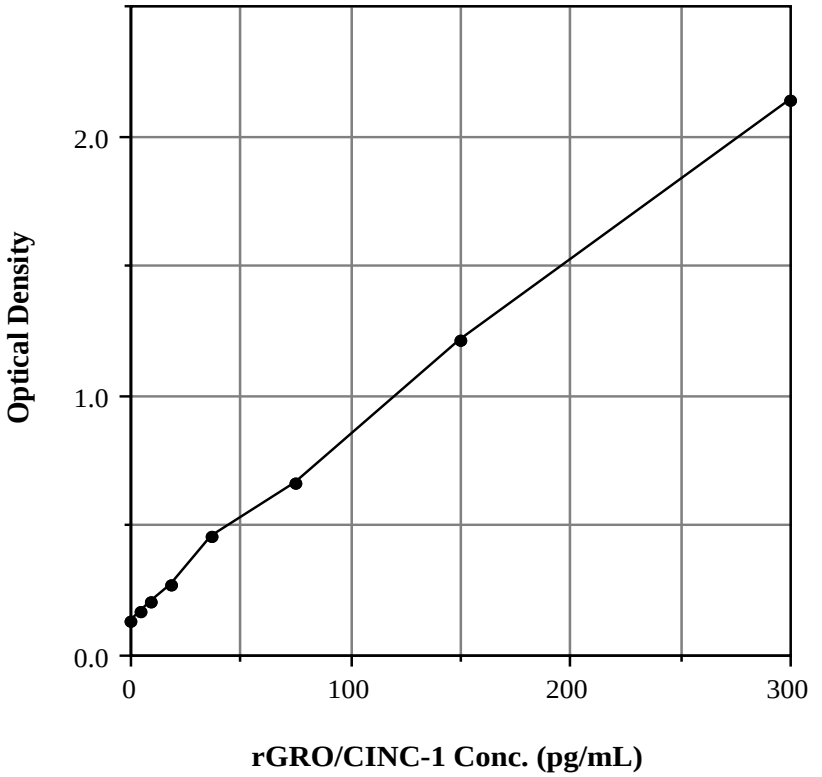
Typical Results

The results shown below are for illustration only and **should not** be used to calculate results from another assay.

Sample	Average OD	Net OD	**rat GRO/CINC-1 (pg/mL)**
Blank	(0.058)		
S0	0.193	0.135	**0**
S1	2.191	2.133	**300**
S2	1.272	1.214	**150**
S3	0.721	0.663	**75**
S4	0.512	0.454	**37.5**
S5	0.333	0.275	**18.8**
S6	0.259	0.201	**9.38**
S7	0.227	0.169	**4.69**

Typical Standard Curve

A typical standard curve is shown below. This curve **must not** be used to calculate rat GRO/CINC-1 concentrations; each user must run a standard curve for each assay.



Performance Characteristics

The following parameters for this kit were determined using the guidelines listed in the National Committee for Clinical Laboratory Standards (NCCLS) Evaluation Protocols[8].

Sensitivity

Sensitivity was calculated by determining the average optical density bound for eight (8) wells run with 0 pg/mL Standard, and comparing to the average optical density for eight (8) wells run with Standard #7. The detection limit was determined as the concentration of rat GRO/CINC-1 measured at two (2) standard deviations from the 0 pg/mL Standard along the standard curve.

Average Optical Density for the S0 =	0.064 ± 0.007 (10.9%)
Average Optical Density for Standard #7 =	0.097 ± 0.007 (7.4%)
Delta Optical Density (4.69-0 pg/mL) =	0.033
2 SD's of the 0 pg/mL Standard = 2 x 0.007 =	0.014

Sensitivity = $\frac{0.014}{0.033}$ x 4.69 pg/mL = **1.99 pg/mL**

Linearity

A sample containing 600 pg/mL rat GRO/CINC-1 was diluted 4 times 1:2 into Assay Buffer and measured in the assay. The data was plotted graphically as actual rat GRO/CINC-1 concentration versus measured rat GRO/CINC-1 concentration.

The line obtained had a slope of 1.085 and a correlation coefficient of 0.999.

Precision

Intra-assay precision was determined by taking samples containing low, medium and high concentrations of rat GRO/CINC-1 and running these samples multiple times (n=24) in the same assay. Inter-assay precision was determined by measuring three samples with low, medium and high concentrations of rat GRO/CINC-1 in multiple assays (n=8).

The precision numbers listed below represent the percent coefficient of variation for the concentrations of rat GRO/CINC-1 determined in these assays as calculated by a 4 parameter logistic curve fitting program.

	rat GRO/CINC-1 (pg/mL)	Intra-assay %CV	Inter-assay %CV
Low	24.19	4.5	
Medium	55.82	2.9	
High	169.47	3.1	
Low	25.05		3.7
Medium	57.34		3.0
High	170.12		3.6

Cross Reactivities

The cross reactivities for a number of related compounds was determined by dissolving the cross reactant in Assay Buffer. These samples were then measured in the rat GRO/CINC-1 assay, and the measured rat GRO/CINC-1 concentration calculated. The % cross reactivity was calculated by comparison with the actual concentration of cross reactant in the sample and expressed as a percentage.

Compound	Cross Reactivity
Rat GRO/CINC-1	100%
Rat GRO/CINC-2α	<0.1%
Rat GRO/CINC-2β	<0.1%
Rat GRO/CINC-3	<0.1%
Rat MCP-1	<0.1%
Rat Rantes	<0.1%
Rat MIP-1	<0.1%
Rat IL-1β	<0.1%

Sample Recoveries

Please refer to pages 4 and 5 for Sample Handling recommendations and Standard preparation.

Rat GRO/CINC-1 concentrations were measured in tissue culture media and rat serum. Rat GRO/CINC-1 was spiked into the undiluted samples of these media which were then diluted with the kit Assay Buffer and assayed in the kit. The following results were obtained:

Sample	% Recovery*	Recommended Dilution*
Tissue Culture Media	104.8	1:4
rat Serum	84.3	1:4

* See Sample Handling instructions on page 4 for details.

References

1. T. Chard, in "An Intro. to Radioimmunoassay & Related Tech.", (1990), 4th Ed., Elsevier, Amsterdam.
2. P. Tijssen, in "Practice & Theory of Enz. Immunoassays", (1985), Elsevier, Amsterdam.
3. Watanabe K, et al., J. Biol. Chem., (1989), 264, 19559-19563.
4. Yamasaki Y, et al., Brain Resarch, (1997), 759 ,103-111.
5. Hashimoto S, et al., Am. J. Physiol., (1996), 270, L819-L828.
6. Tsuruma T, et al., Transplant Proc., (1998), 30, 2644-2645.
7. Makita H, et al., Am. J. Respir. Crit. Care Med., (1998), 158, 573-579.
8. NCCLS Evaluation Protocols, SC1, (1989), NCCLS, Villanova, PA, 19085.

LIMITED WARRANTY

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Material Safety Data Sheet (MSDS) available on our website or by fax.

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